

Lentiviral-Mediated Gene Therapy (RP-L102) for Fanconi Anemia [Group A] is Associated with Polyclonal Integration Patterns in the Absence of Conditioning

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Philip Ancliff, MA, MRCP, MRCPATH; Elena Sebastián MD, PhD; Michael Rothe, PhD; Antonella L Bastone, PhD;
Philipp John-Neek, MSc.; Axel Schambach, PhD; Wei Wang, PhD; Ilana Moscatelli, PhD; Grace Choi, BS;
Miriam Zeini, PhD; Eileen Nicoletti, MD; Cindy Eide, MS; John E Wagner MD; Gayatri R Rao, MD, JD;
Adrian Thrasher, MBBS, PhD; Jonathan D Schwartz, MD; Maria G Roncarolo, MD; Juan A Bueren, PhD

27th Annual Meeting of the American Society of Gene and Cell Therapy (ASGCT)

May 10, 2024

Abstract # 245

Disclosures

- Consultant: 48 Bio, GV, Lyrik Therapeutics, Prime Medicine, Spotlight Therapeutics, Stemodontics, Teiko Bio
- Current Equity: 48 Bio, Beam Therapeutics, Decibel Therapeutics, Editas Medicine, GV, Lyrik Therapeutics, Magenta Therapeutics, Prime Medicine, Spotlight Therapeutics, Stemodontics, Teiko Bio, Divested Equity in last 24 months: Forty Seven, Inc., Global Blood Therapeutics
- Intellectual property rights: Gilead Sciences, Jasper Therapeutics, Magenta Therapeutics
- Research funding from Rocket Pharmaceuticals, Inc. Jasper Pharmaceuticals, STRM.Bio

Introduction

Fanconi anemia (FA) is a rare inherited disorder of defective deoxyribonucleic acid (DNA) repair characterized by:¹⁻⁴

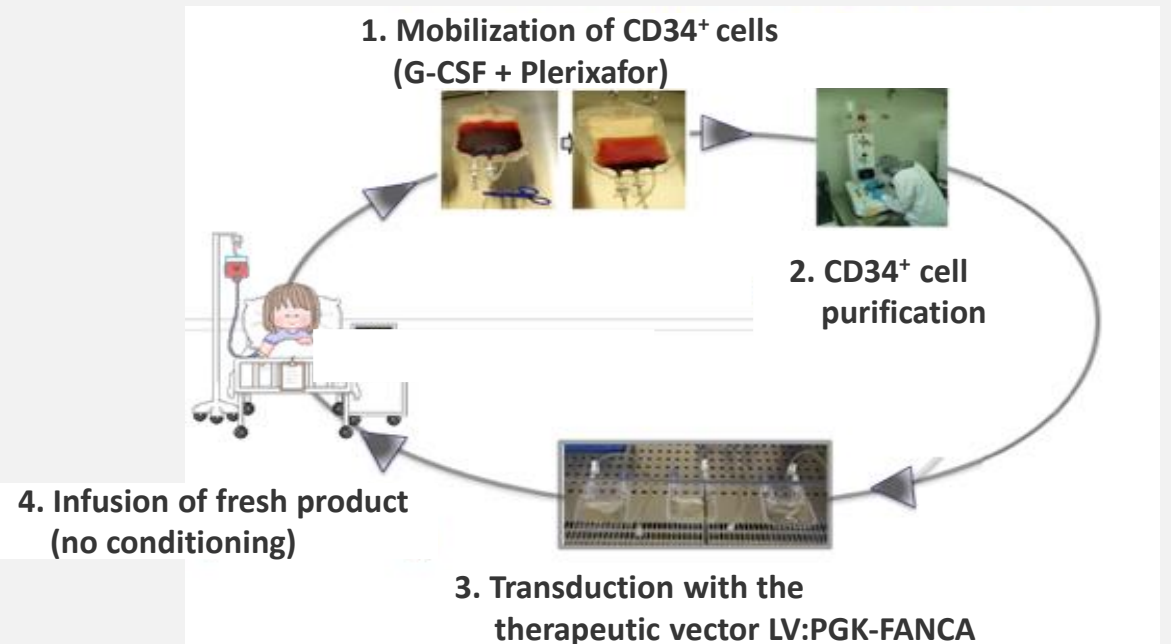
- Progressive bone marrow failure (BMF)
- Cancer Predisposition
- Congenital abnormalities

FA represents a significant unmet medical need¹⁻⁴

- FA-A accounts for ~70% of FA
- Allogeneic HSCT is potentially curative of the hematologic component but confers significant toxicity
 - 100-day transplant-associated mortality
 - GvHD
 - 3-4× increased risk for solid organ malignancies
 - Long-term coronary artery disease, endocrinopathies & neurocognitive dysfunction

RP-L102 Rationale

- Insertion of a functional *FANCA* gene into autologous FA-A CD34+ cells confers resistance to DNA-damage & provides proliferative advantage to modified cells enabling engraftment without cytotoxic conditioning⁵



GvHD=graft versus host disease; LV=lentiviral vector.

1. Anur P et al. *Bone Marrow Transplant*. 2016; 51(7): 938-944; 2. Ebens C et al. *Biol Blood Marrow Transplant*. 2018; 24(4):765-771; 3. Guardiola P et al. *Blood*. 2004;103(1): 73-77; 4. Mehta P et al. *Blood*. 2017; 129(16): 2308-2315; 5. Río P et al. *Nat Med* 2019; 25:1396-1401.

Global RP-L102 Studies

Key Eligibility Criteria

Inclusion:

- FA complementation group A
- Minimum age: 1y
- BM CD34+ cell concentration: $\geq 30/\mu\text{L}$ (from aspirate)

Exclusion:

- Available & eligible HLA-identical sibling donor
- MDS or leukemia (including associated cytogenetic abnormalities)
- Mosaicism with stable/improved blood counts

Endpoints

Efficacy:

Phenotypic correction: Increased resistance of BM progenitor cells (CFCs) to DNA-damaging agent mitomycin-C (MMC)

Engraftment: Peripheral blood (PB) and BM vector copy number (VCN)

Clinical response: Hematologic stabilization

Safety: Evaluation of safety and tolerability

Demographics and Investigational Product Metrics

n=14 patients

Patient #14	Age at Enrollment (years)	Follow Up (months)	CD34+ Cell Dose (cells/kg)	CFCs/kg	Mean VCN:		Transduction Efficiency in CFCs (%)	CFC Survival MMC 10nM (%)
					Liquid Culture	CFCs		
1 (1001)	5	54	2.0×10^5	5.2×10^4	2.08	0.62*	67	33
2 (1002)	6	18 [†]	3.7×10^5	5.0×10^4	2.21	0.92*	72	47
3 (2004)	3	42	4.8×10^5	1.3×10^5	1.70	0.73	100	63
4 (2008)	2	36	3.2×10^6	5.5×10^5	1.65	1.56	97	63
5 (2009)	3	28 [‡]	1.9×10^6	3.1×10^5	2.16	0.76	61	45
6 (2010)	3	36	4.1×10^6	n/a	0.62	n/a	n/a	n/a
7 (2011)	5	36	2.8×10^6	n/a	1.46	n/a	n/a	n/a
8 (2014)	6	32	5.4×10^5	3.6×10^4	3.68	1.93	87	31
9 (2016)	2	32	3.0×10^5	2.5×10^4	1.96	0.64	88	64
10 (2021)	2	21	2.3×10^6	4.3×10^5	1.55	1.97	78	38
11 (2023)	5	18	2.5×10^5	2.8×10^4	1.70	2.16	87	50
12 (2024)	1	15	1.8×10^6	1.7×10^5	1.69	1.91	88	93
13 (2026)	1	4	1.9×10^6	1.2×10^5	1.30	0.80	75	51
14 (2025)	3	2	3.5×10^5	5.6×10^4	3.45	2.50	92	63

All patients ≤ 6 years old at enrollment

12 patients have ≥ 12 months of follow-up

Trial enrollment is complete

Median Values

VCN (liquid culture): 1.70
VCN (CFC): 1.75
Transduction efficiency: 87.0%
CFC MMC-resistance: 50.5%

*Mean CFC VCN was assessed from a cryopreserved drug product sample.

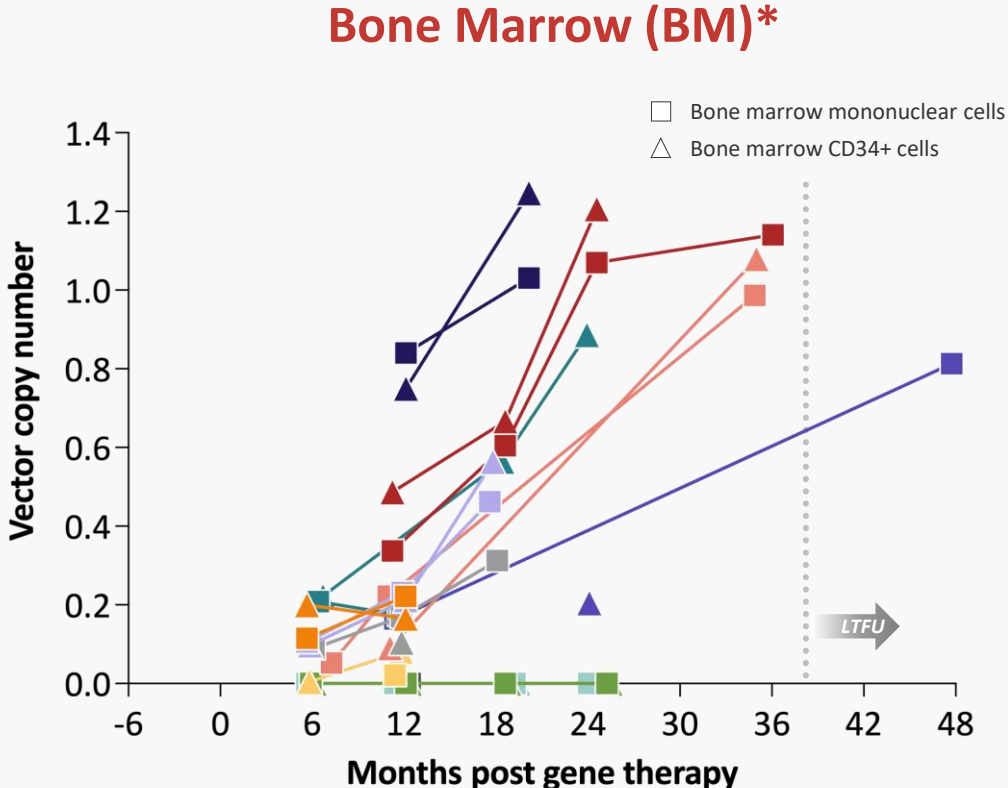
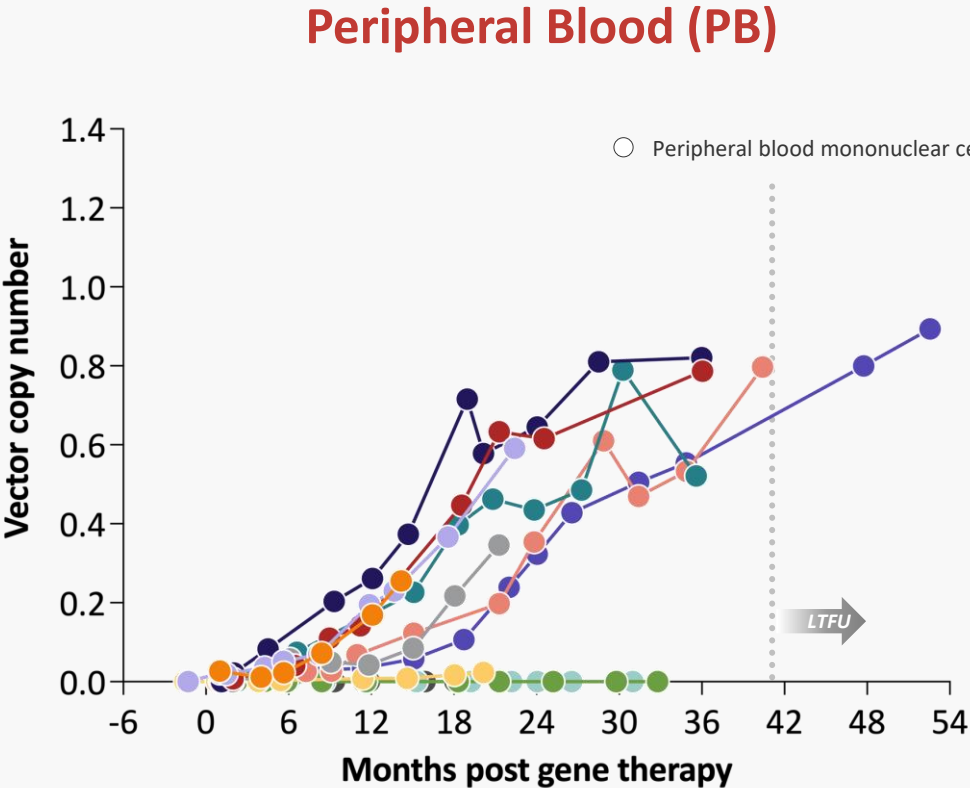
†Patient withdrawn from RP-L102 study at 18 months post-RP-L102 infusion; received successful allogeneic HSCT for BMF. Safety follow up continues on LTFU study.

‡Patient withdrawn from RP-L102 study at 28 months post-RP-L102 infusion; received successful allogeneic HSCT for NHL. Safety follow up continues on LTFU study.

BMF=Bone marrow failure; CFC=colony-forming cell; HSCT=Hematopoietic stem cell transplantation; LTFU=long-term follow-up; MMC=Mitomycin-C; n/a: not available; NHL=Non-Hodgkin Lymphoma; VCN=vector copy number.

Data cutoff: September 11, 2023

Progressively Increasing and Sustained Genetic Correction in 8 of 12 Patients at ≥ 1 year post-RP-L102



- 1 (1001)
- 2 (1002)
- 3 (2004)
- 4 (2008)
- 5 (2009)
- 6 (2010)
- 7 (2011)
- 8 (2014)
- 9 (2016)
- 10 (2021)
- 11 (2023)
- 12 (2024)

Progressive gene marking increases in PB and BM associated with hematologic stability

*Vector copy number in bone marrow not available at some stipulated time points due to insufficient sample to perform assay.
LTFU=long-term follow-up.

Insertion Site Analysis (ISA) Methodology

- ISA was performed using INSPIRED (INtegration Site Pipeline for paired-End readDs)
- Samples[‡] analyzed for:
 - abundance of individual integration sites over time
 - diversity/polyclonality of samples over time
 - presence of common integration sites
 - proximity of integrations to proto-oncogenes

Gini index: Describes how equally clones contribute to IS pool.

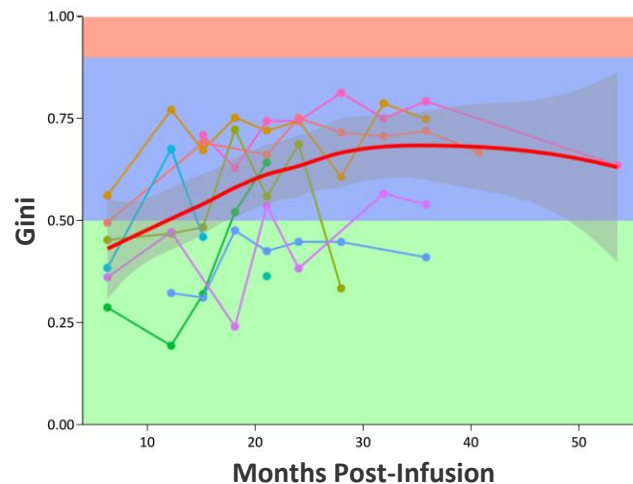
Low value: polyclonal insertion pattern and even sequence distribution

High value: unequal contribution of more dominant sequences to the overall sequence pool

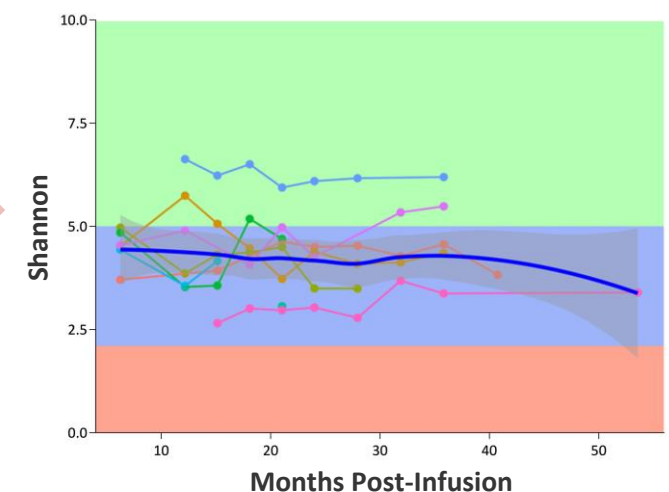
Shannon index: Describes the overall diversity of the IS pool

- ≤ 2.1 : oligoclonal
- > 2.1 and ≤ 5.0 : polyclonal
- > 5.0 : highly polyclonal

Gini Index across Phase 2 Studies*



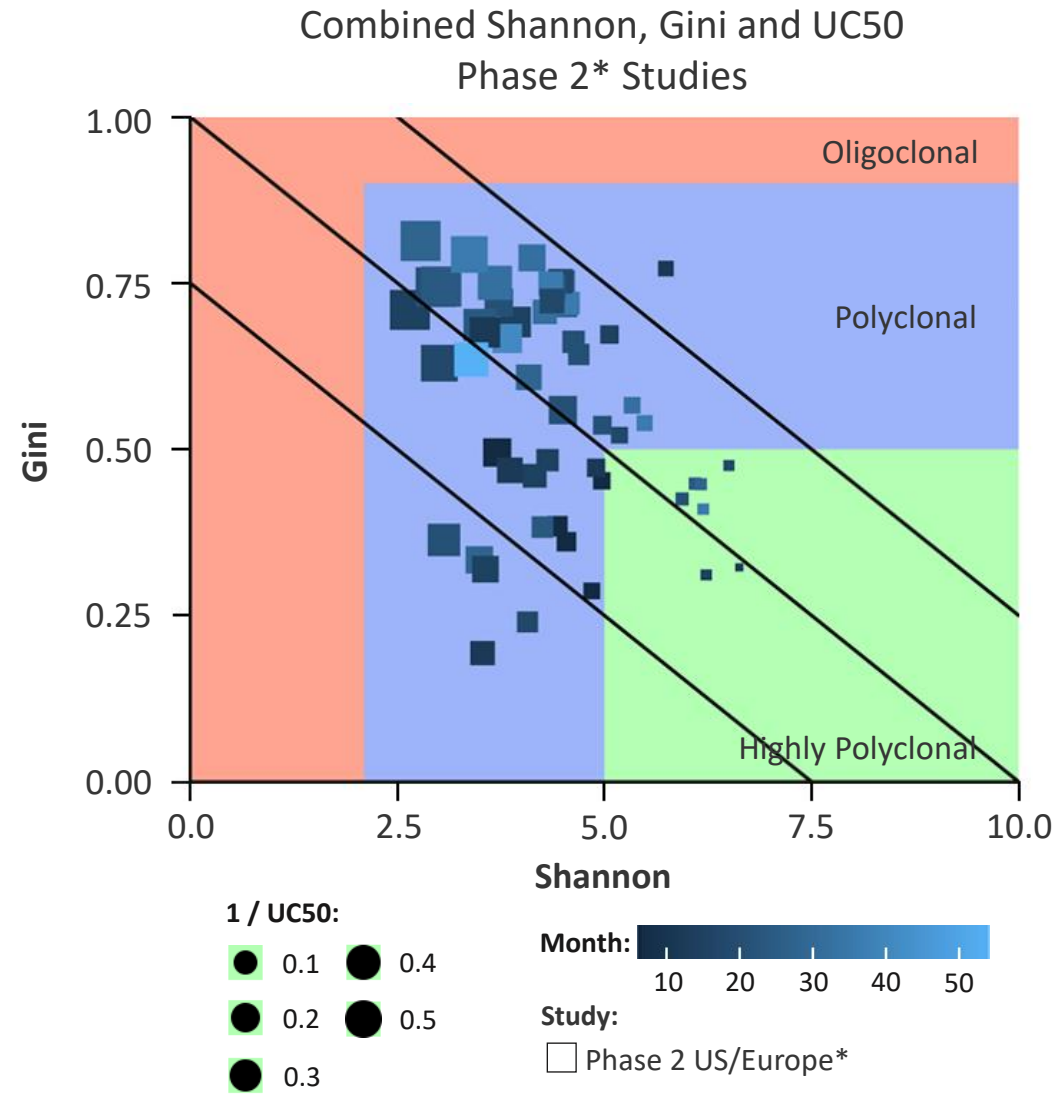
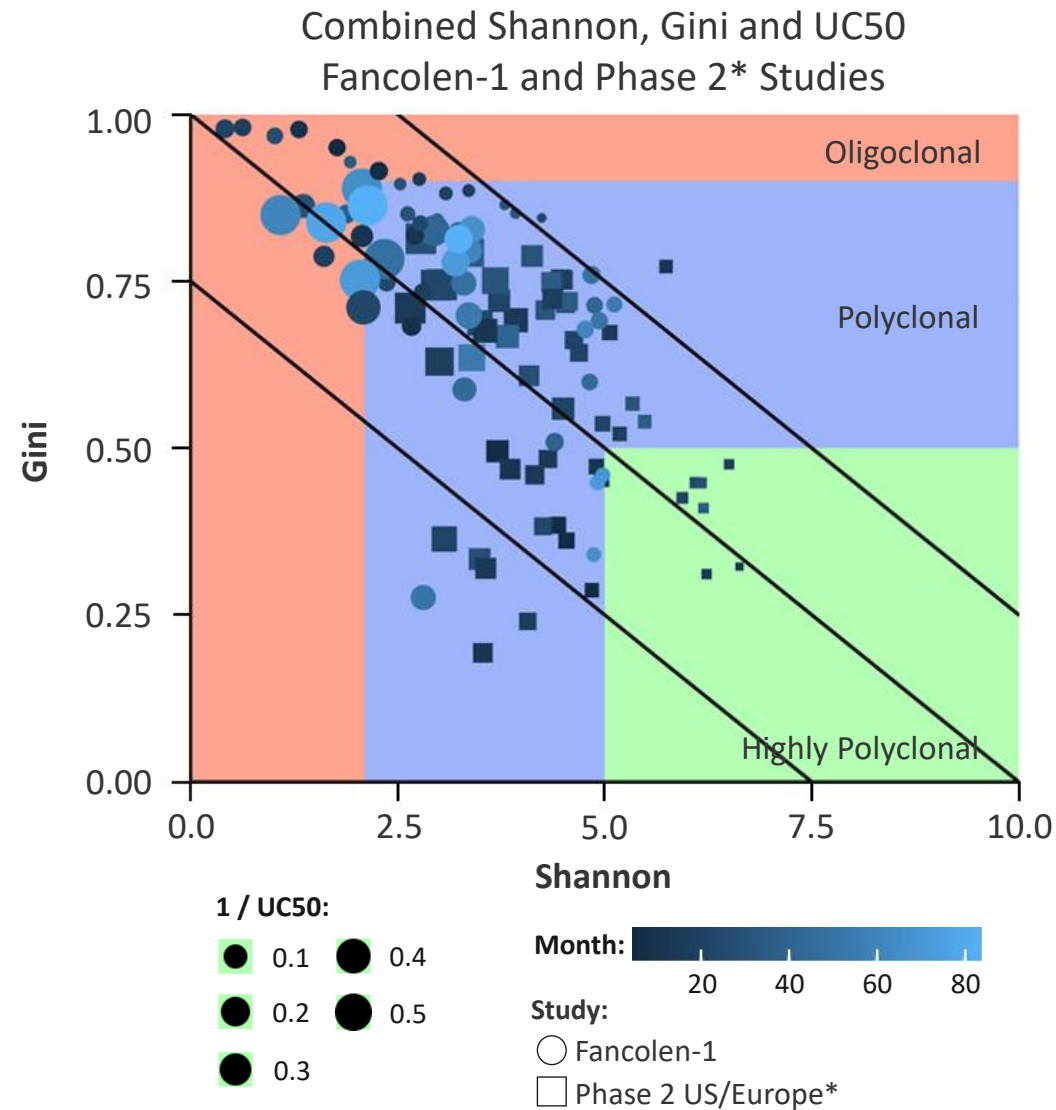
Shannon Index across Phase 2 Studies*



[‡] ISA evaluated if VCN ≥ 0.02

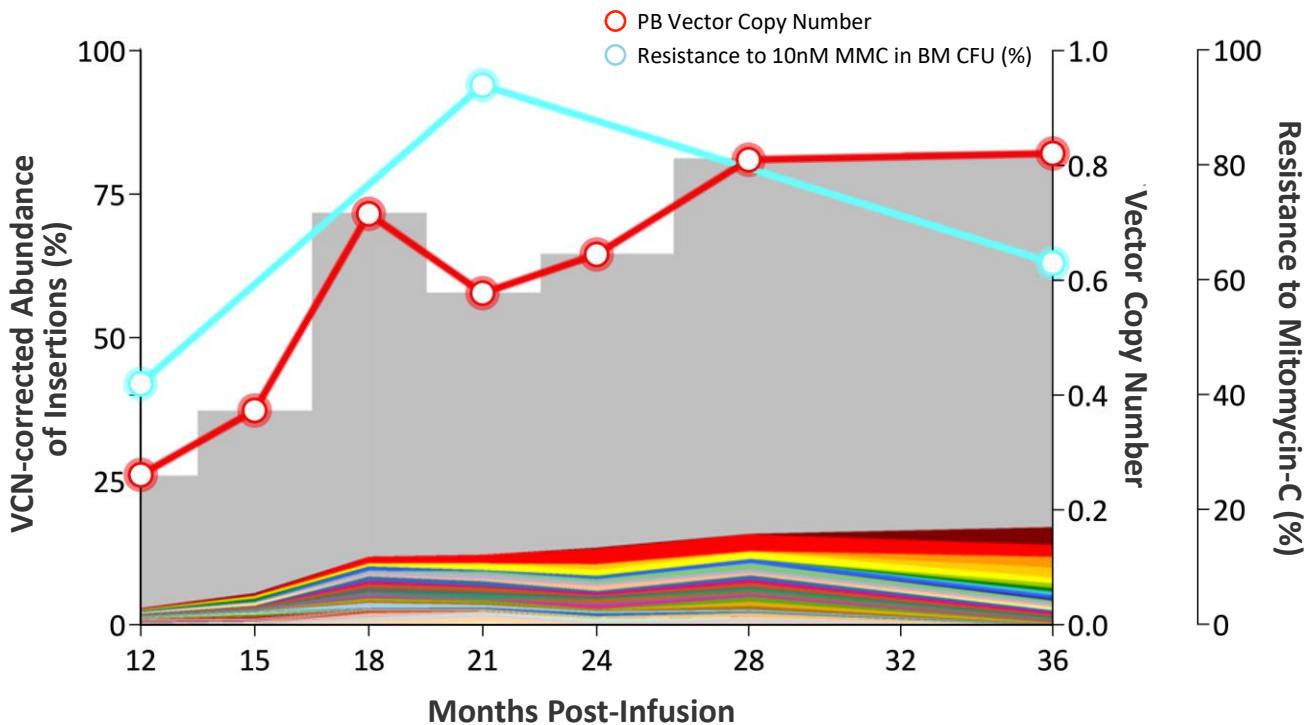
* Phase 2 studies include RP-L102 sponsored US Phase 1 & 2 and EU/UK Phase 2 (Fancolen-2).
IS=integration site.

Aggregate Diversity Indexes for Fancolen-1 and Phase 2 Studies



*Phase 2 studies include RP-L102 sponsored US Phase 1 & 2 and EU/UK Phase 2 (Fancolen-2).
UC50= describes how many integrations in a sample contribute to 50% of all sequences

Representative Longitudinal ISA: Patient 6 (2010)



- Colored areas represent all integrations detected in top 10 integrations at any timepoint
- The gray area represents all other identified integrations and reflects the VCN (in %) at each timepoint
- Tables below display the VCN-corrected contributions of top 10 integrations at each timepoint

VCN-Corrected Contributions of Top 10 Integrations

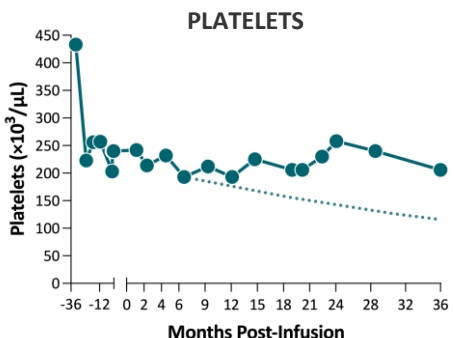
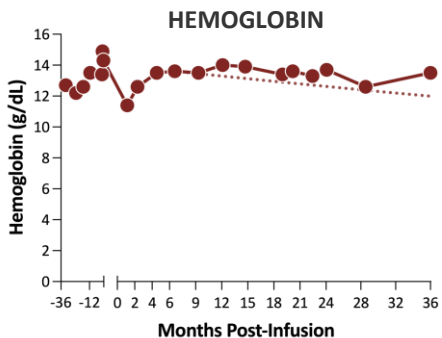
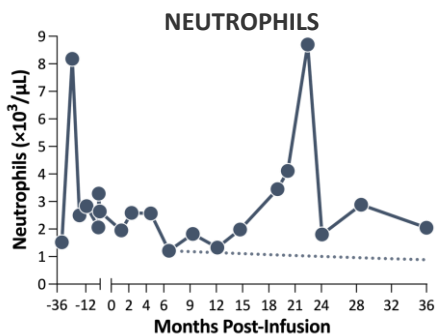
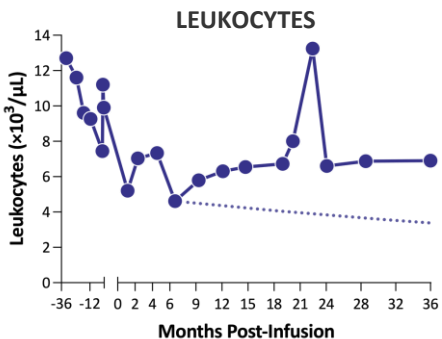
Month 12	Month 15	Month 18	Month 21	Month 24	Month 28	Month 36
0.25%: KDM2A*	0.42%: TPX2*	1.11%: NR2C2*	1.49%: NR2C2*	2.66%: NR2C2*	2.89%: NR2C2*	3.10%: CBFβ~*
0.21%: LOC105371046*	0.40%: BRD4~*	0.94%: FAT4*	1.18%: TLE3	0.97%: NLF3*	1.24%: TLE3	2.06%: NR2C2*
0.18%: DYRK1A~*	0.40%: ILF3*	0.83%: TCF20*	1.00%: VPS13C*	0.94%: TLE3	1.08%: THCAT158*	1.89%: CNOT6L*
0.17%: DOCK7*	0.33%: CBFβ~*	0.73%: CYSLTR1	0.83%: BRD4~*	0.85%: CARD8~*	0.87%: SEC16A*	1.73%: NLF3*
0.15%: CD33*	0.33%: GOLGA2*	0.71%: POLK*	0.83%: TCF20*	0.73%: SEMA3C	0.74%: POLK*	1.01%: TLE3
0.15%: CNM2*	0.30%: VAV1~*	0.71%: VPS13C*	0.76%: ARHGAP35*	0.70%: PPP6R2*	0.74%: TCF20*	0.78%: SPN53*
0.14%: MYO9B*	0.30%: VPS13C*	0.69%: PACS1*	0.66%: ZDHHC17*	0.61%: THCAT158*	0.74%: ZNF318~*	0.65%: ILF3*
0.13%: FAT4*	0.26%: ARHGAP35*	0.60%: BRD4~*	0.62%: SEC14L1*	0.59%: SNAP23*	0.67%: ARHGAP35*	0.59%: VSTM5
0.13%: NR2C2*	0.26%: NLF3*	0.56%: ARHGAP35*	0.52%: THCAT158*	0.53%: BRD4~*	0.67%: ARHGAP35*	0.52%: POLK*
0.13%: POLE*	0.23%: DYRK1A~*	0.54%: WASF2~*	0.45%: CYSLTR1	0.53%: GFI1B~*	0.67%: C6orf15	0.46%: DHRS2

* Integration within a transcription unit
~ Integration within 50kb of a proto-oncogene

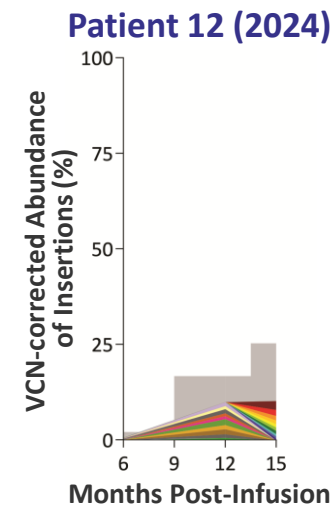
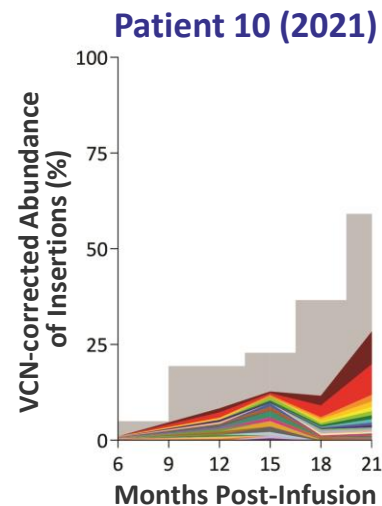
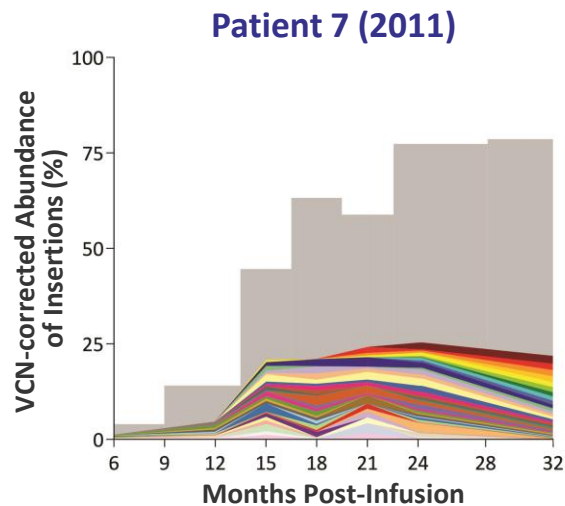
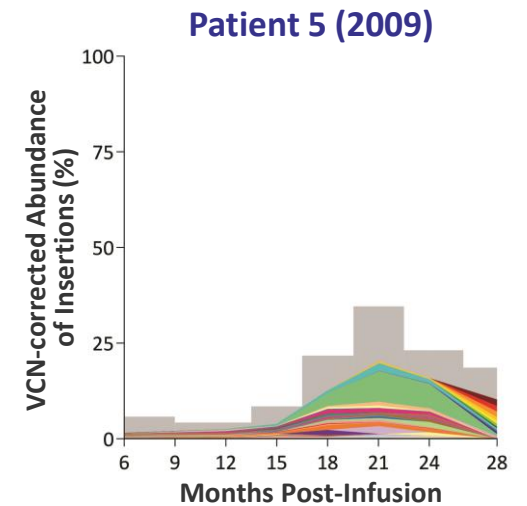
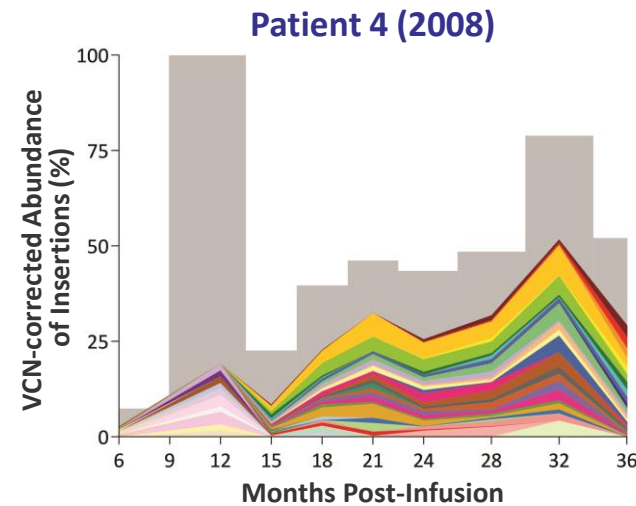
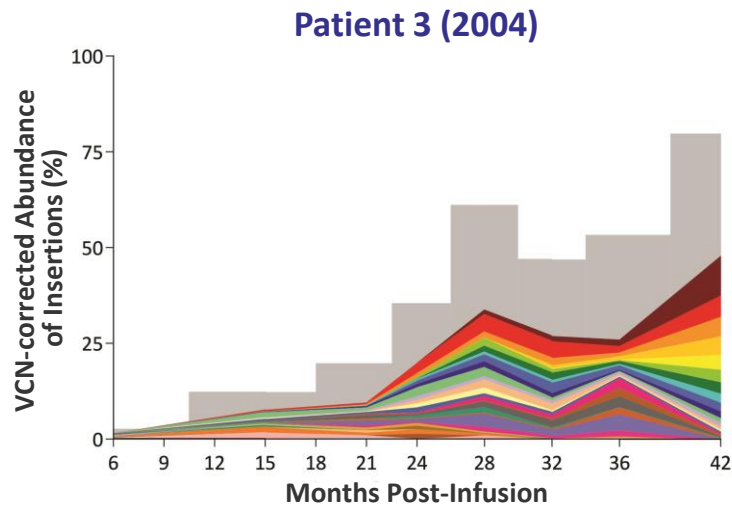
%VCN-Corrected Abundance of Insertions = [% contribution to all UIS] × [VCN] if PB VCN < 1.0 OR
= [% contribution to all UIS] × [1.0] if PB VCN ≥ 1.0

BM=bone marrow; BMF=Bone marrow failure; CFU=Colony-forming unit; MMC=Mitomycin-C; PB=Peripheral blood;
RBC=Red blood cell; VCN: Vector copy number; UIS=Unique insertion sites.

Blood count graphs: Dotted lines indicate projected blood count decreases based on natural history evaluation from n=139 FA-A patients (IFAR registry, data on file)



Longitudinal ISA in Additional Patients with Evidence of Comprehensive Engraftment

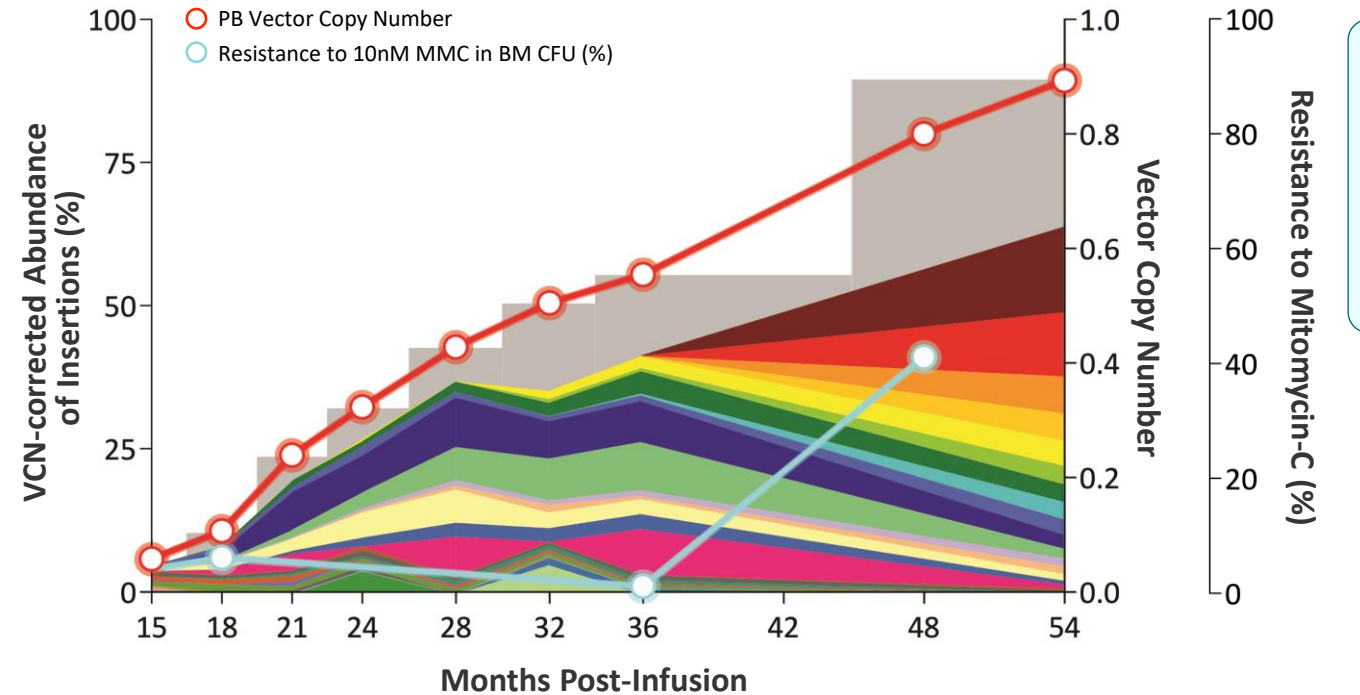


ISA=insertion site analysis; MMC=Mitomycin-C; VCN=vector copy number.

Data cutoff date: September 11, 2023

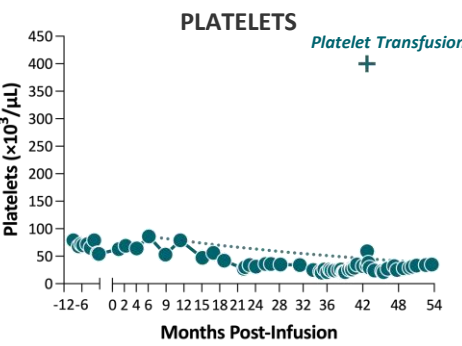
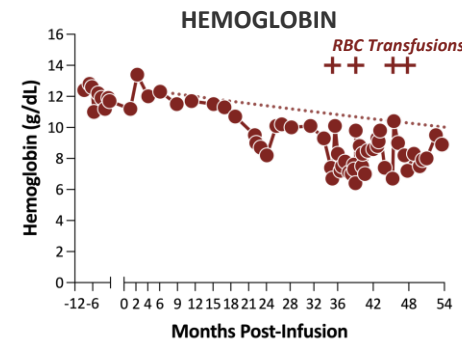
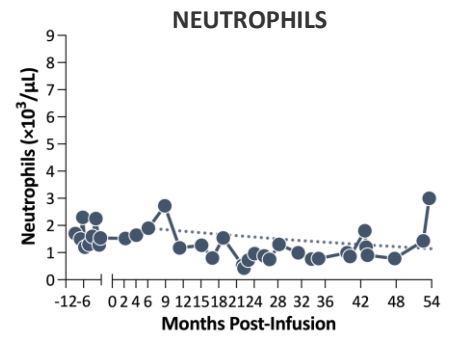
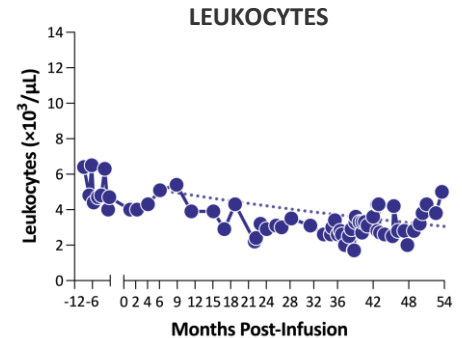
Less Robust Polyclonality Associated with More Limited Engraftment: Patient 1 (1001)

(Including blood count declines during initial 24-months post-infusion)



- Following initial decline requiring transfusion support, blood counts have stabilized, concurrent with the development of BM MMC resistance and sustained genetic correction
- Blood counts remain above thresholds for severe BMF, with no transfusion requirements in most recent 6 months

Blood count graphs: Dotted lines indicate projected blood count decreases based on natural history evaluation from n=139 FA-A patients (IFAR registry, data on file)



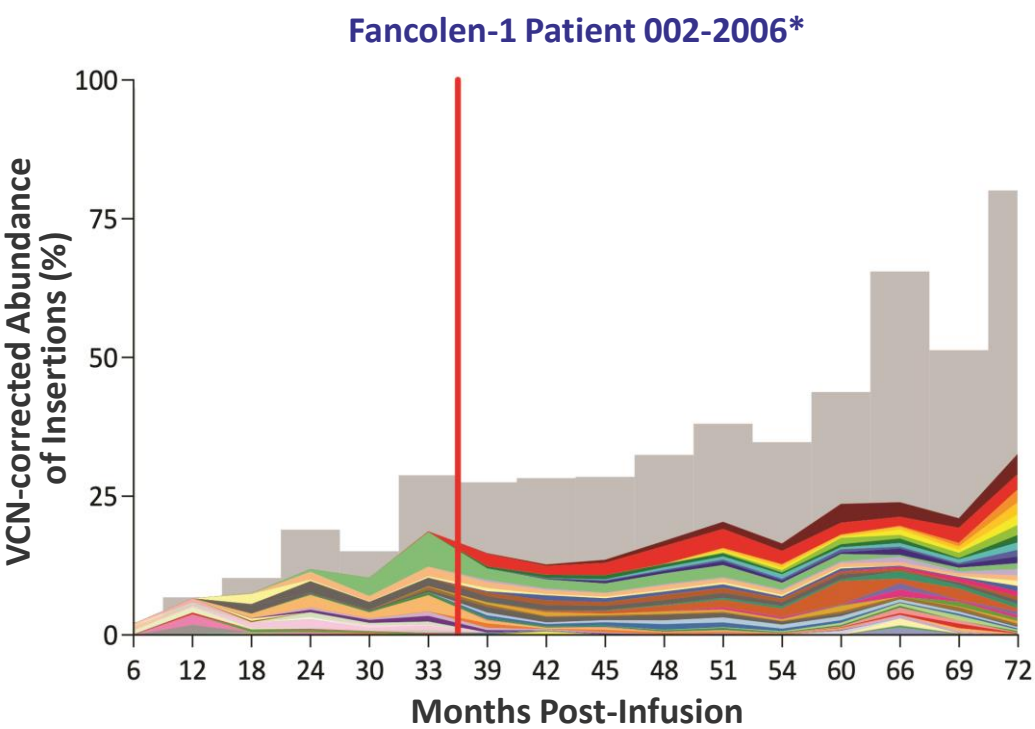
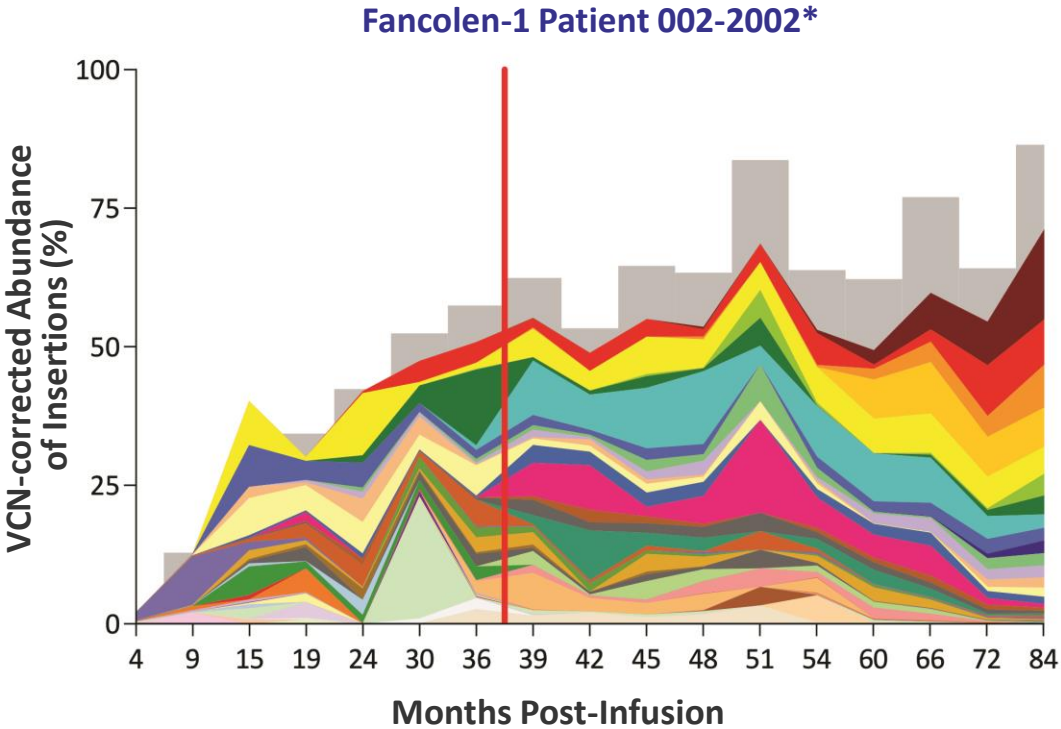
VCN-Corrected Contributions of Top 10 Integrations							
Month 15	Month 18	Month 21	Month 24	Month 28	Month 32	Month 36	Month 54
1.78%: POU2F1*	1.78%: CNOT6L*	6.55%: CNOT6L*	6.36%: CNOT6L*	8.57%: CNOT6L*	7.23%: SLC8A1	8.35%: SLC8A1	14.86%: WDR82*
1.34%: CARD8~*	1.34%: CARD8~*	2.89%: POU2F1*	4.41%: EGR2	6.90%: POU2F1*	6.46%: CNOT6L*	7.89%: POU2F1*	11.15%: SLC1A6*
1.17%: POU5F1P3*	1.17%: EGR2	2.14%: EGR2	4.04%: POU2F1*	5.80%: EGR2	5.08%: POU2F1*	7.09%: CNOT6L*	6.42%: HNRNP1*
0.89%: LINC00571	0.89%: POU2F1*	1.26%: SLC8A1	2.73%: SLC8A1	5.69%: SLC8A1	2.76%: EGR2	3.86%: FCHSD2*	4.73%: TLK1*
0.82%: CNOT6L*	0.82%: LINC00571	1.14%: FCHSD2*	1.68%: SKAP1*	2.42%: LRMDA*	2.32%: LRMDA*	2.63%: EGR2	4.39%: FERMT3*
0.75%: EGR2	0.75%: SKAP1*	0.97%: LINC00571	1.50%: LRMDA*	1.75%: FCHSD2*	2.28%: FCHSD2*	2.60%: LRMDA*	3.21%: RCC2~*
0.45%: ATF1~*	0.45%: POU5F1P3*	0.84%: SKAP1*	1.14%: CARD8~*	1.01%: SKAP1*	1.46%: FERMT3*	2.09%: FERMT3*	3.04%: FCHSD2*
0.24%: CHM*	0.24%: EYA3*	0.81%: ASH1L*	0.95%: POU5F1P3*	0.99%: LY86	1.33%: MSH5*	1.00%: LY86	3.04%: LINC02639
0.21%: INTS6~*	0.21%: CNOT6L*	0.71%: CARD8~*	0.82%: FCHSD2*	0.82%: CARD8~*	1.29%: OAZ1	1.00%: SKAP1*	2.70%: SKAP1*
0.21%: SKAP1*	0.21%: NSRP1*	0.52%: GFI1B~	0.73%: GFI1B~	0.73%: CNOT6L*	0.86%: SKAP1*	0.94%: OAZ1	2.36%: CNOT6L*

* Integration within a transcription unit
~ Integration within 50kb of a proto-oncogene

%VCN-Corrected Abundance of Insertions = [% contribution to all UIS] × [VCN] if PB VCN <1.0 OR
= [% contribution to all UIS] × [1.0] if PB VCN ≥1.0

BM=Bone marrow; BMF=Bone marrow failure; CFU=Colony-forming unit; MMC=Mitomycin-C; PB=Peripheral blood; RBC=Red blood cell; VCN: Vector copy number; UIS=Unique insertion sites.

Polyclonal ISA Patterns in Fancolen-1 Patients Receiving Higher Cell Doses and/or VCNs with Hematologic Stability and Increases Over 6-7 Years



*ISA strata to left of red bars indicate retrospective sample analysis at U Hannover; strata to right of bar were performed prospectively.
ISA=insertion site analysis; VCN=vector copy number.

Conclusions

- RP-L102 is a potentially curative therapy to prevent FA-related BMF, which can be administered without a suitable allogeneic donor or transplant related toxicities
 - 8 of 12 patients with ≥ 1 year of follow up have demonstrated sustained and progressively increasing genetic correction
 - Genetic correction associated with phenotypic correction and hematologic stability
- RP-L102 has conferred polyclonal insertion patterns in Phase 2 studies (utilizing Process B manufacturing optimizations)
 - Relative to Fancolon-1, Phase 2 patients demonstrated higher polyclonality and diversity of gene modified cells
 - Overall 1.3-fold higher mean VCN
 - 1.9-fold higher number of unique integrations
 - 1.4-fold lower Gini index
 - 1.5 fold higher Shannon index
- These diverse integration patterns are indicative of long-term hematopoietic stem cell repopulation of the BM and PB and clonal diversity in the absence of conditioning, without evidence of clonal dominance or potential mutagenesis

Acknowledgements



Julián Sevilla, MD, PhD

Josune Zubicaray, MD, PhD

Elena Sebastian MD, PhD



Juan Bueren, PhD

Paula Río, PhD

Susana Navarro, PhD



Lucile Packard
Children's Hospital
Stanford



Rajni Agarwal, MD

Maria Grazia Roncarolo, MD, PhD



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Adrian J. Thrasher, MBBS, PhD, FMedSci

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Grainne O'Toole, RN

Jinhua Xu-Bayford, RN



John E. Wagner, MD

Margaret MacMillan, MD

Cindy Eide, MS



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Antonella Lucía Bastone, PhD

Philipp John-Neek, M.Sc.



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Ilana Moscatelli, PhD

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